



Predictive markers of lung cancer – recent updates

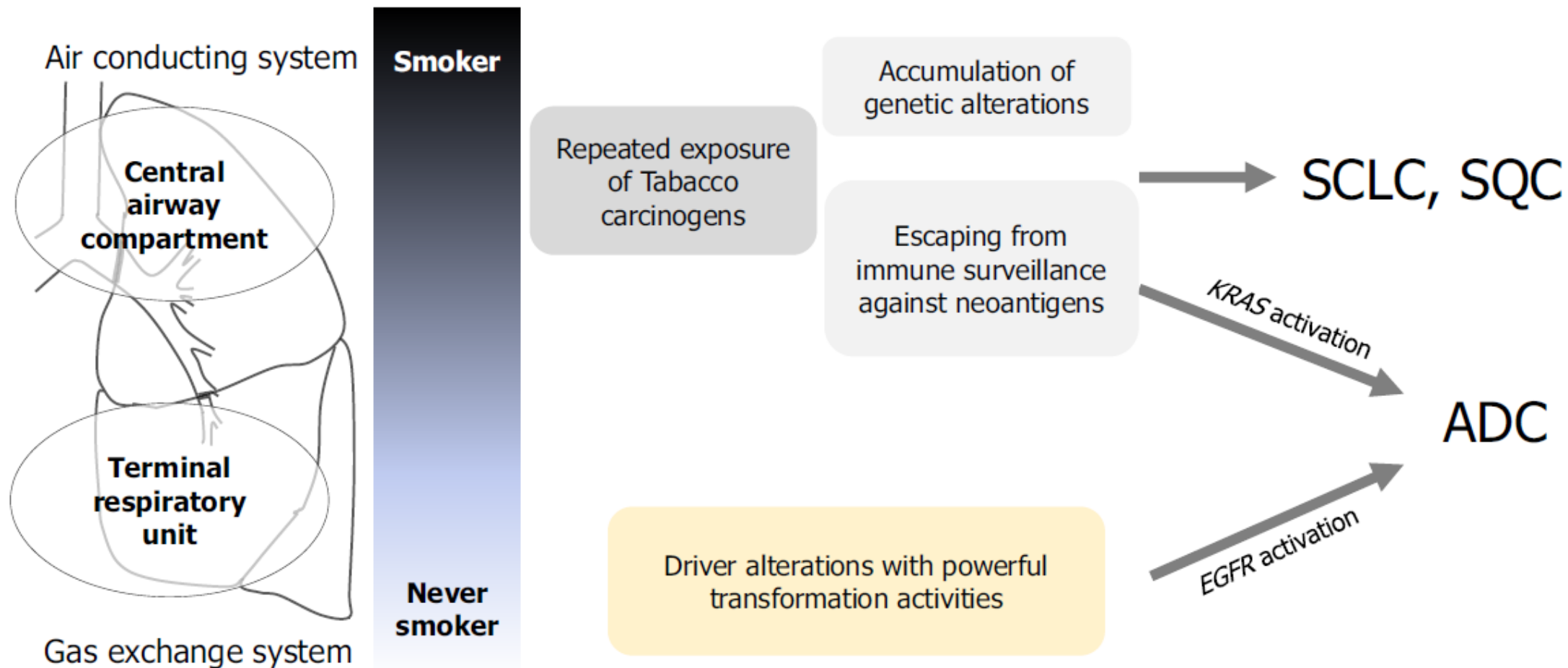
Izidor Kern

University Clinic Golnik, Slovenia

agenda

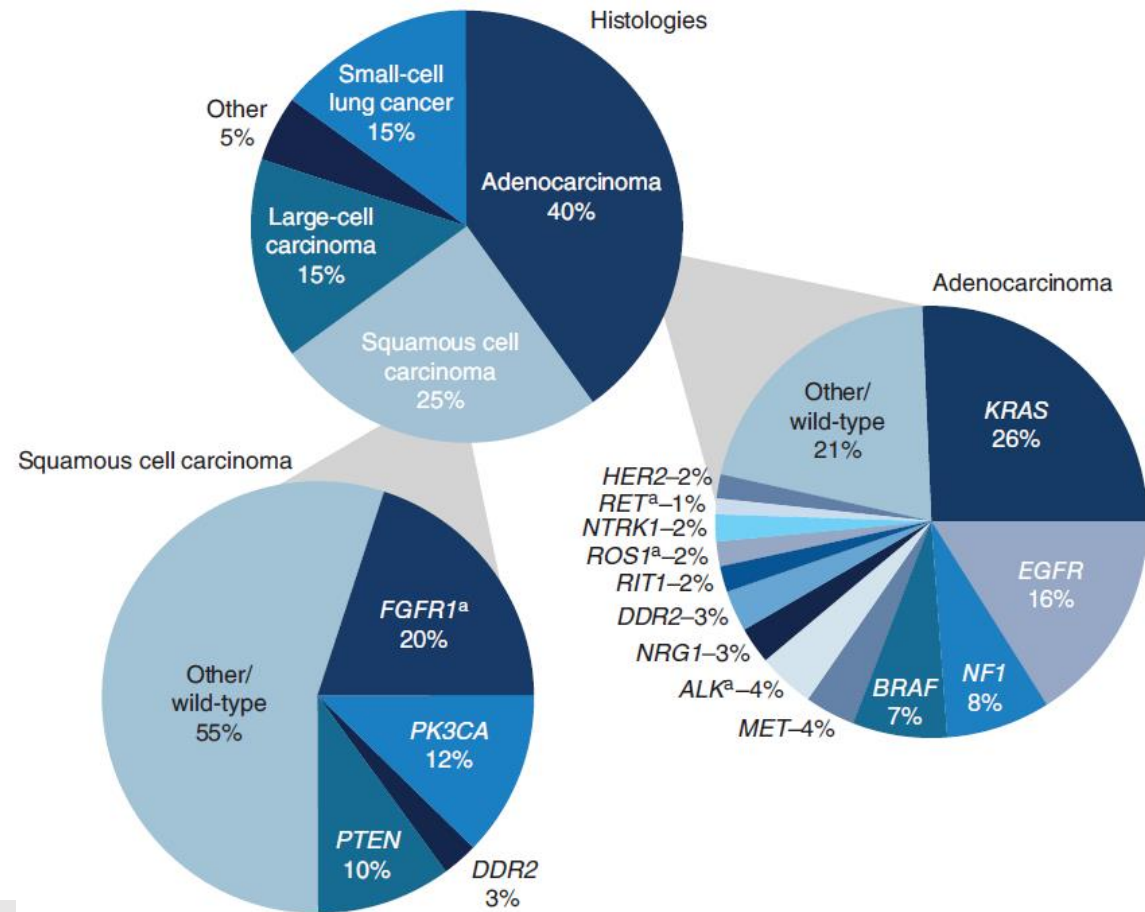
1. NSCLC genetic abnormalities and driver mutations
2. Molecular biomarker testing
Methods, early stages, emerging biomarkers, resistance mechanisms
3. PD-L1
4. Liquid biopsy
5. Multiple lung tumours
6. Testing implementation and obstacles

Lung adenocarcinoma is not one disease

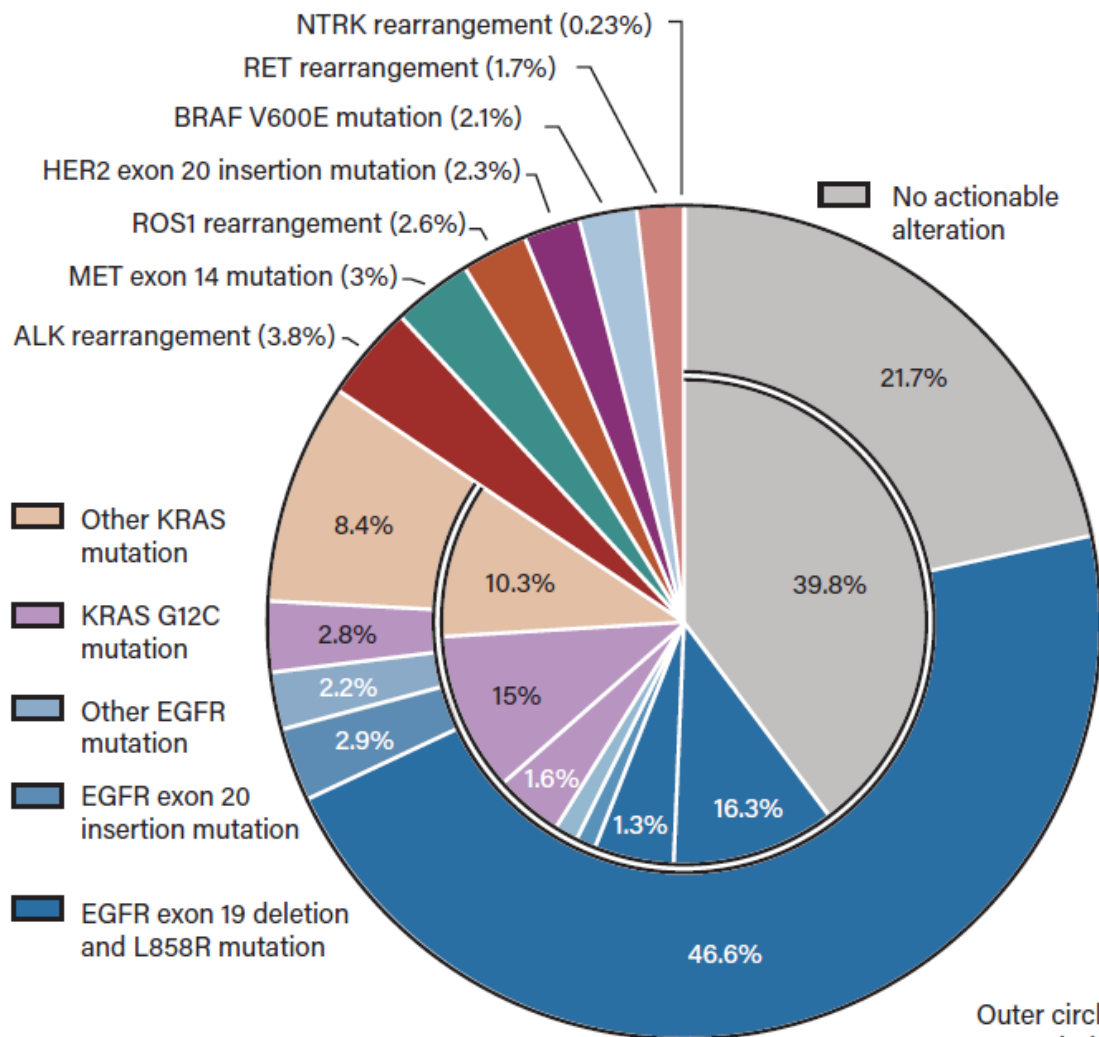


NSCLC and molecular drivers

Oncogenes	Tumor-suppressor genes	
<i>AKT1</i>	<i>KIT</i>	<i>BRCA2</i>
<i>AKT2</i>	<i>KRAS</i>	<i>CDKN2A</i>
<i>ALK</i>	<i>MET</i>	<i>KEAP1</i>
<i>BRAF</i>	<i>NRAS</i>	<i>MLL2</i>
<i>DDR2</i>	<i>NRF2</i>	<i>NF1</i>
<i>EGFR</i>	<i>NTRK1/2/3</i>	<i>PTEN</i>
<i>ERBB2</i>	<i>PIK3CA</i>	<i>RB1</i>
<i>FGFR1</i>	<i>RET</i>	<i>STK11</i>
<i>HRAS</i>	<i>RICTOR</i>	<i>TP53</i>
<i>JAK3</i>	<i>ROS1</i>	<i>TSC1</i>
	<i>SOX2</i>	<i>VHL1</i>



NSCLC oncogenic drivers



Tan AC, J Clin Oncol 2022

Litomyšl 2025

EGFR

Exo 19 deletion
L858R point mutation in exon 21
L861Q point mutation in exon 21
G719X point mutation in exon 18
T790M point mutation exon 20

NSCLC: $\cong 15\%$ in Western populations
NSCLC: $\cong 35-50\%$ in Asian populations
LUAD: 27%
LSQCC: $<9\%$
LUAD never smoker: 42%
LUAD former smoker: 13.5%
LUAD current smoker: 5%

ALK

ALK rearrangement
EML4-ALK (the most frequent)
KIF5B-ALK
KLC1-ALK

NSCLC: 3-5%
LUAD: 3-7%
LSQCC: 0.2-1%
LUAD never smoker: 5-11%
LUAD smoker: 0-0.8%

ROS1

ROS1 rearrangement
CD74-ROS1
SDC4-ROS1
SLC34A2-ROS1
EZR-ROS1

NSCLC: 0.5-2%
LUAD: 2.6%
LSQCC: 0%
LUAD never smoker: 5.8%
LUAD smoker: 0.4%

RET

RET rearrangement

NSCLC: 1-2%
LUAD: 1.4%
LSQCC: 0%
LUAD never-smoker: 1.8%
LUAD smoker: 0.7%

BRAF

Point mutations
BRAF^{V600E} (30-40% of cases)
BRAF^{nonV600E} (60-70% of cases):
BRAF^{V469A}, BRAF^{D594G},
BRAF^{G466A}
Class I, II and III BRAF mutants

NSCLC: 2.6%
LUAD: 1.4%
LSQCC: 0.2-1%
LUAD never-smoker: 5-11%
LUAD smoker: 0-0.8%

HER2

Point mutations
HER2 amplifications

NSCLC: 1.8%
LUAD: 2.3%
LSQCC: 0%
LUAD never smoker: 4.5%
LUAD smoker: 0.8%

MET

MET mutations (exon 14 skipping mutations)
MET amplifications
MET translocations

Met mutations, METex 14
NSCLC: 2.5-3%
LUAD: 3%
LSQCC: 0%
LUAD never smoker: 4%
LUAD smoker: 2.2%

LC stage and driver alterations

Driver muta

Tumor type

W

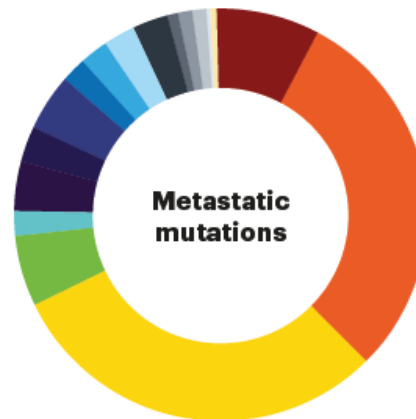
Early-stage ADC
(n=240)

Entire cohort
(n=1,125)

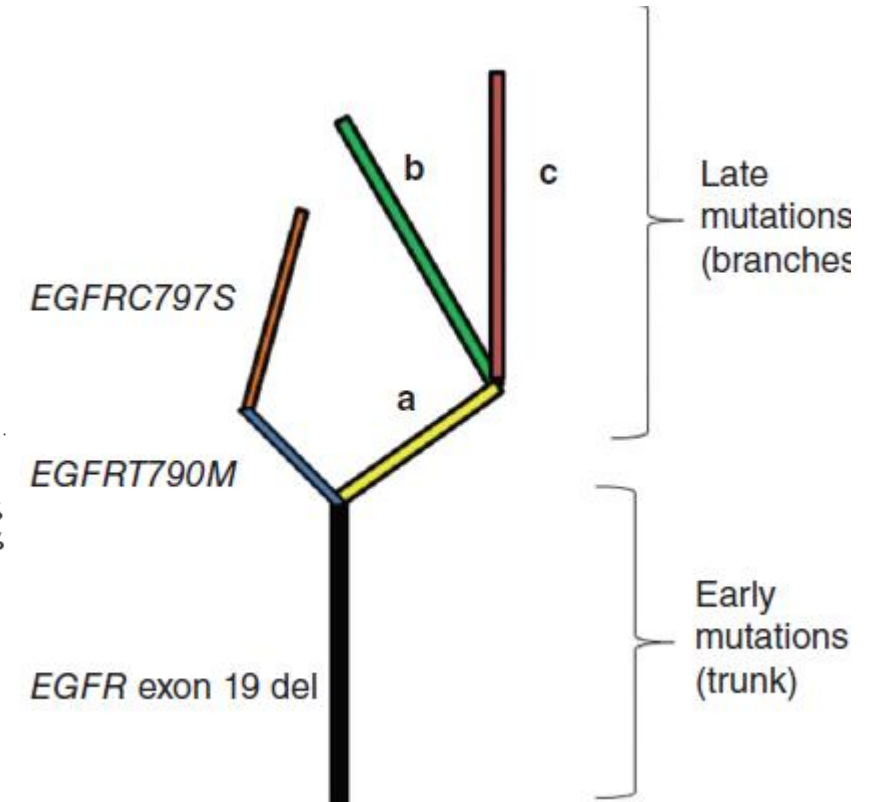
ADC, adenocarcinoma; Early invasive ADC and entire cohort



Other genes 27.3%
KRAS 29.1%
EGFR 14.2%
BRAF 7.2%
NF1 truncation 6.3%
ERBB2 1.8%
MET splice 1.4%
ALK fusion 0.8%
ROS1 fusion 0.9%
RET fusion 0.3%
MET amplification 1.7%
ERBB2 amplification 1.6%
MAP2K1 2.2%
NRAS 0.5%
HRAS 0.5%
RIT1 1.6%
FGFR1 or FGFR2 2.6%

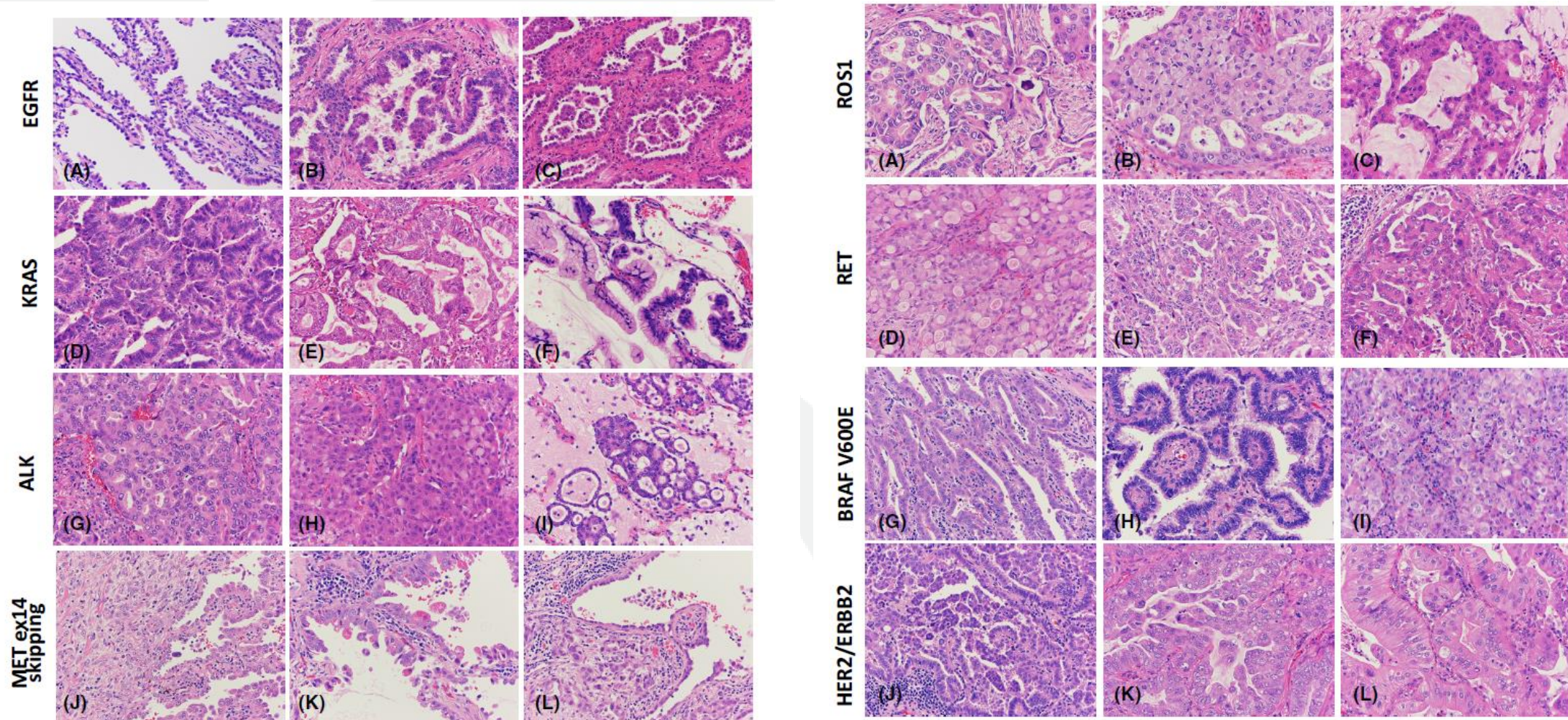


Other genes 7.8%
KRAS 29.9%
EGFR 30.3%
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NF1 truncation 1.9%
ERBB2 3.8%
MET splice 3%
ALK fusion 4.4%
ROS1 fusion 1.9%
RET fusion 2.3%
MET amplification 2.5%
ERBB2 amplification 2.7%
MAP2K1 0.7%
NRAS 1.2%
HRAS 1.2%
RIT1 0.2%
FGFR1 or FGFR2 0.7%

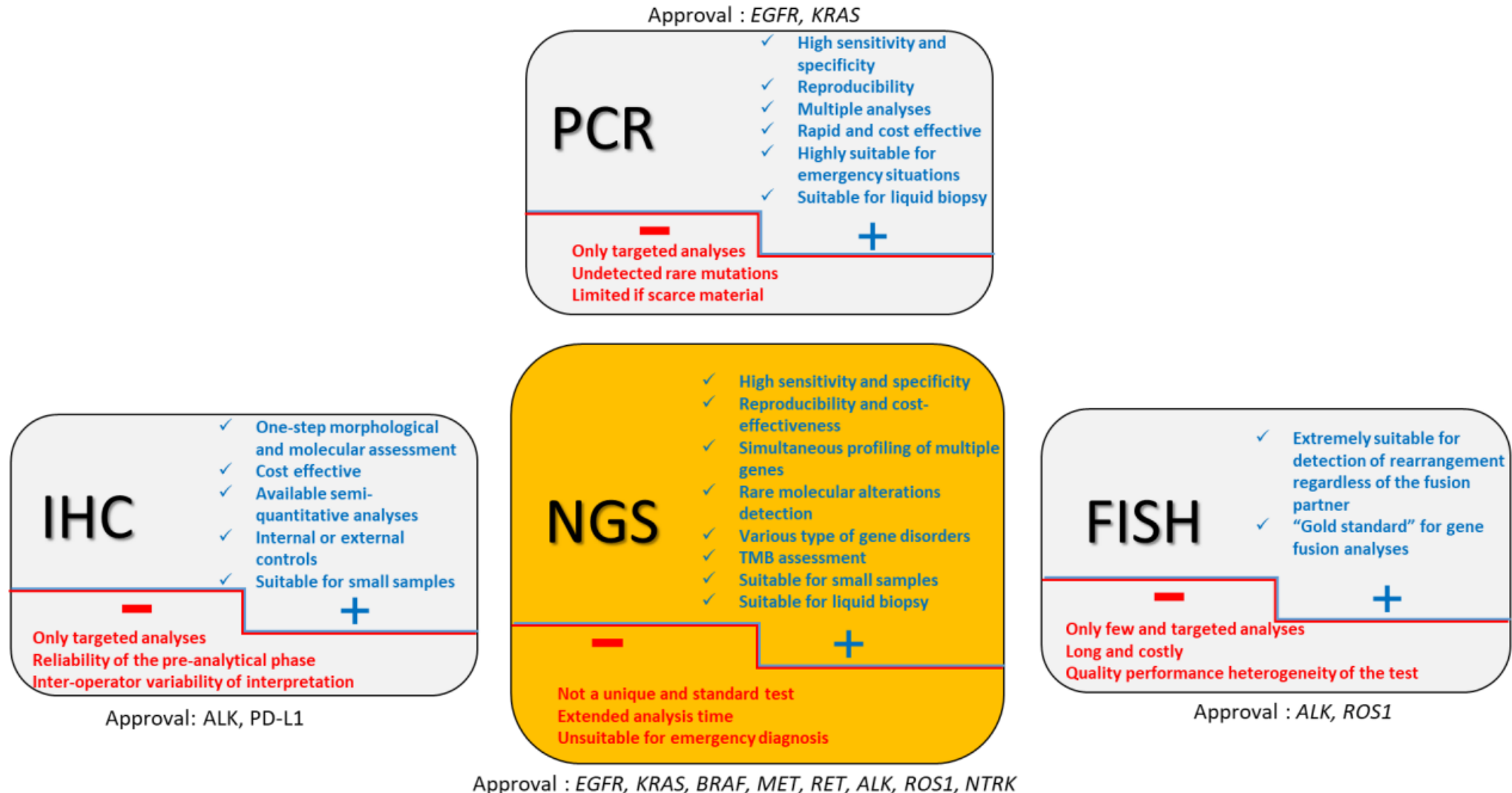


Adenocarcinoma

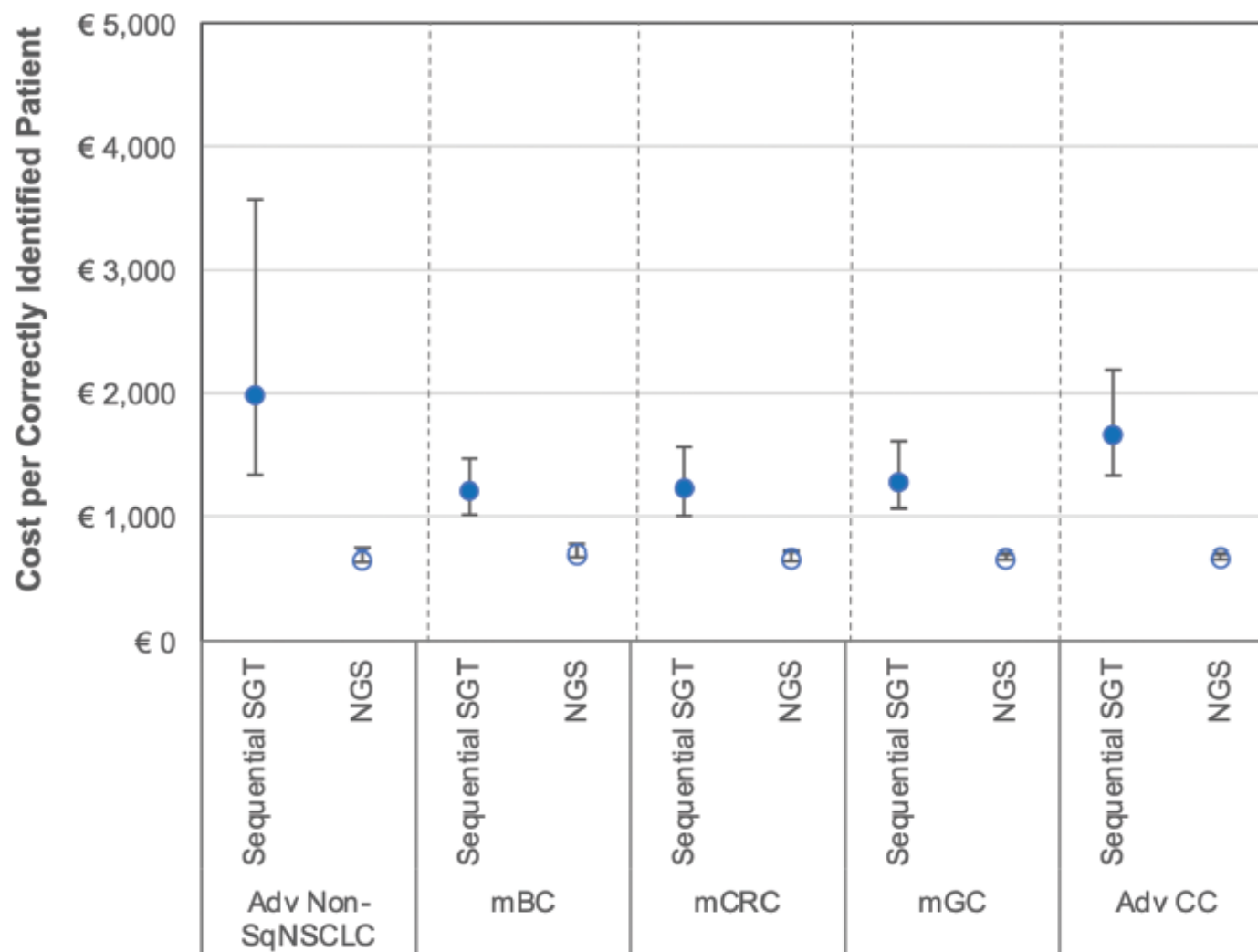
All targetable mutations identified in LC are associated with ADC



Predictive biomarker testing methods



Predictive biomarker testing cost

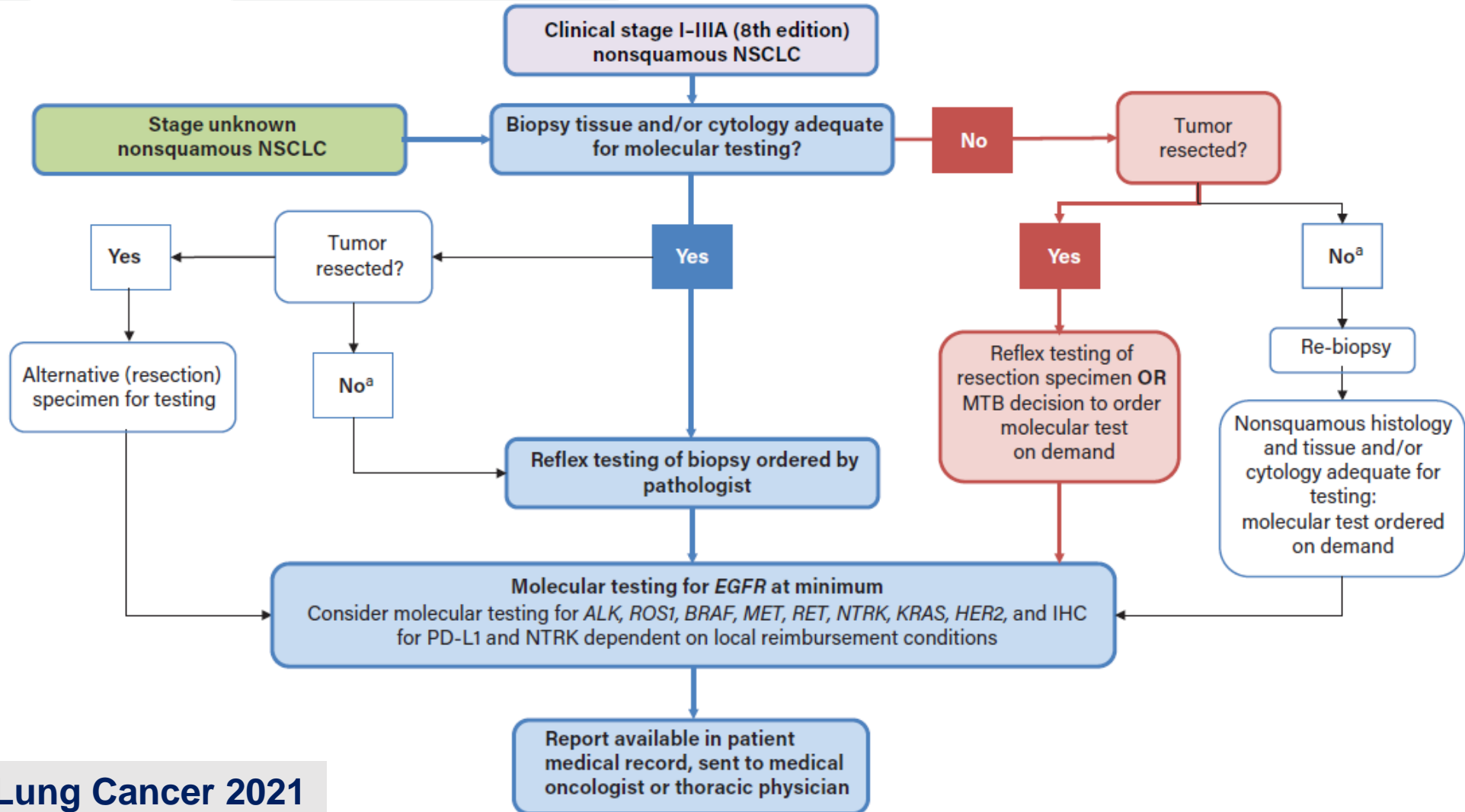


Predictive biomarker testing - status

Biomarker	Austria	Belgium	Czech Republic	England ^{&}	France	Germany	Netherlands	Portugal	Slovenia	Spain	Sweden
<i>EGFR</i>	● 2013	● 2010	● 2012	● n/a	● 2008	● 2012	● 2011	● 2013	● 2010	● 2012	● 2009
<i>ALK</i>	● 2013	● 2013	● 2013	● n/a	● 2012	● 2012	● 2015	● 2013	● 2013	● 2012	● 2013
<i>ROS1</i>	● 2013	● 2017	● 2016	● n/a	● 2012	● 2015	● 2015	●* 2022	● 2015	● 2020	● 2014
<i>BRAF V600</i>	● 2020	● 2017	●*	● n/a	● 2012	● 2016	● 2015	●* 2022	● 2018	● 2020	● 2018
<i>RET</i>	● 2020	● 2022	●*	● n/a	● 2018	● 2016	● 2015	●* 2022	● 2022	● 2023	● 2023
<i>MET</i> exon 14	● 2020	● 2020	●*	● n/a	● 2012	● 2018	● 2020	●* 2022	● 2022	● 2023	● 2023
<i>MET</i> amp	● 2020	● 2023	●	● n/a	● [†] 2023	● [#] 2018	●	●* 2022	●	● 2023	● [§] 2023
<i>KRAS G12C</i>	● 2020	● 2021	●*	● n/a	● 2008	● 2021	● 2015	●* 2022	● 2022	● 2023	● 2022
<i>NTRK</i>	● 2020	● 2021	●*	● n/a	● 2018	● 2016	● 2020	●* 2022	● 2020	● 2023	● 2022
<i>HER2/ERBB2</i>	● 2020	● 2016	●*	●	● 2022	● 2016	● 2015	●* 2022	● 2022	● 2023	●* 2018
<i>NRG1</i>	● 2020	●	●	●	● 2022	● [#] 2019	● 2020	●	●	●	●* 2019
<i>PD-L1</i>	● 2020	● 2018	● 2016	● n/a	● 2015	● 2016	● 2020	●	● 2017	● 2020	● 2015

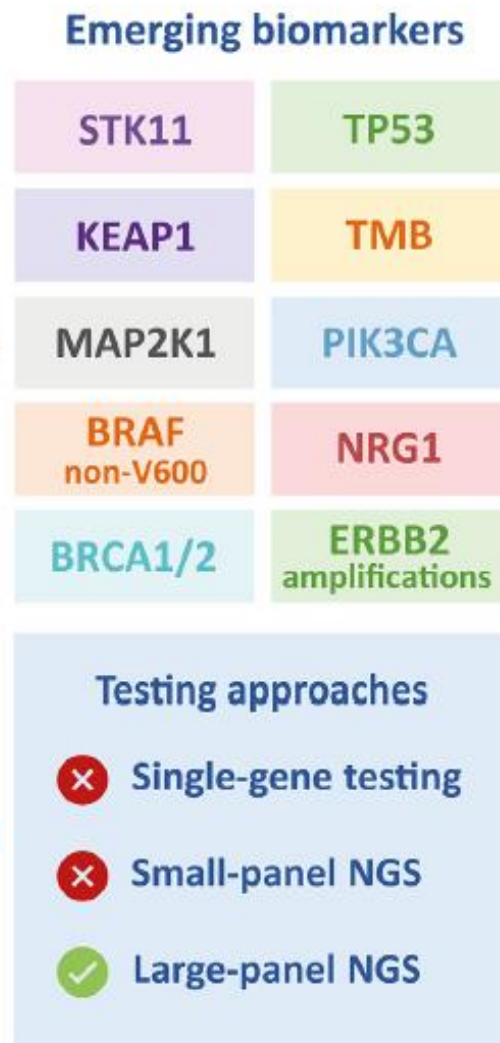
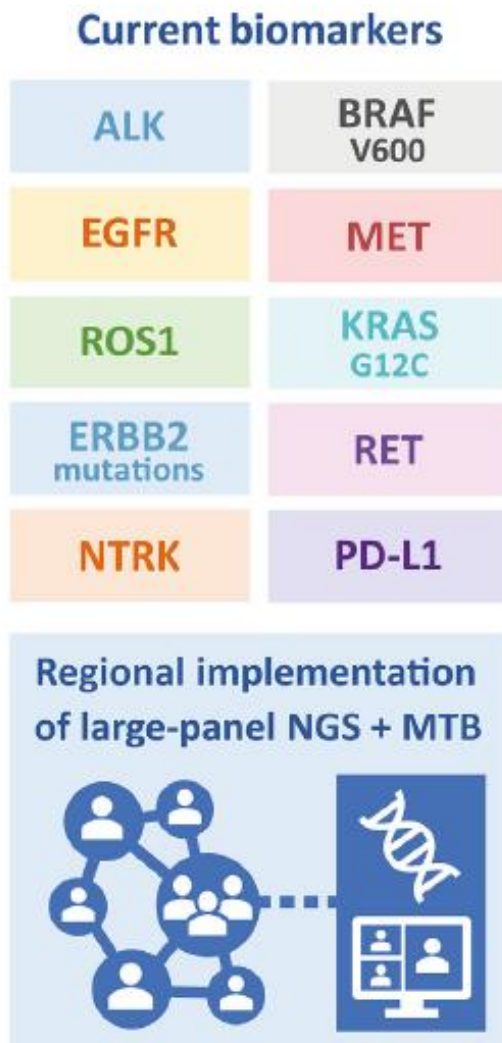
●, required by current national guidelines; ●, recommended by current national guidelines; ●*, not specified in current national guidelines but recommended by national expert consensus; ●, testing not recommended or biomarker not present in current national guidelines.

Reflex testing in early stages



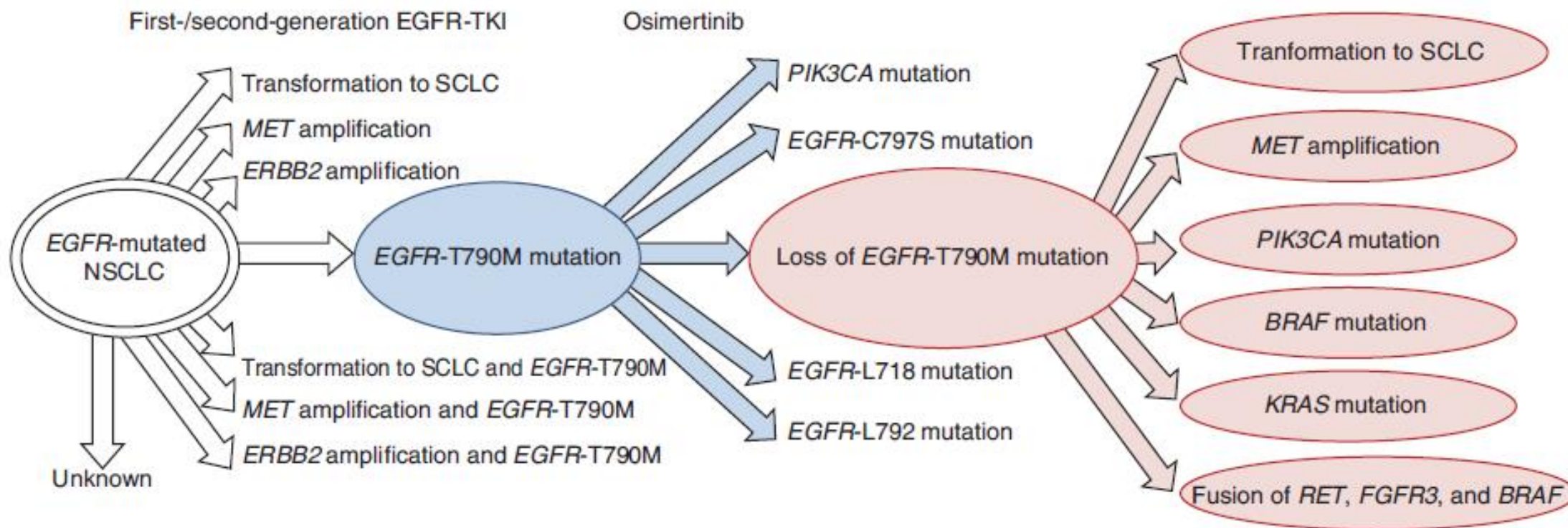
NSCLC – predictive biomarker testing

Mandatory and highly recommended molecular tests		
MANDATORY ESCAT I	ALK	Fusion
	BRAF	V600
	EGFR	Common
	MET	Mutation
	NTRK	Fusion
	RET	Fusion
	ROS1	Fusion
Highly RECOMMENDED ESCAT II	EGFR	Exon 19
	HER2	Mutation
	KRAS	G12C
	MET	Amplification



Mandatory and potential molecular biomarkers	
TP53	Mutation
RB1	Mutation
RBM10	Mutation
AKT	Mutation
CTNNB1	Mutation
JAK2/3	Mutation
NRAS	Mutation
HRAS	Mutation
High TCR clonality	
High CD8 density	
High dNLR/LIPI	
Adequate microbiota	
Gut and tumor DNA, metabolites, products	

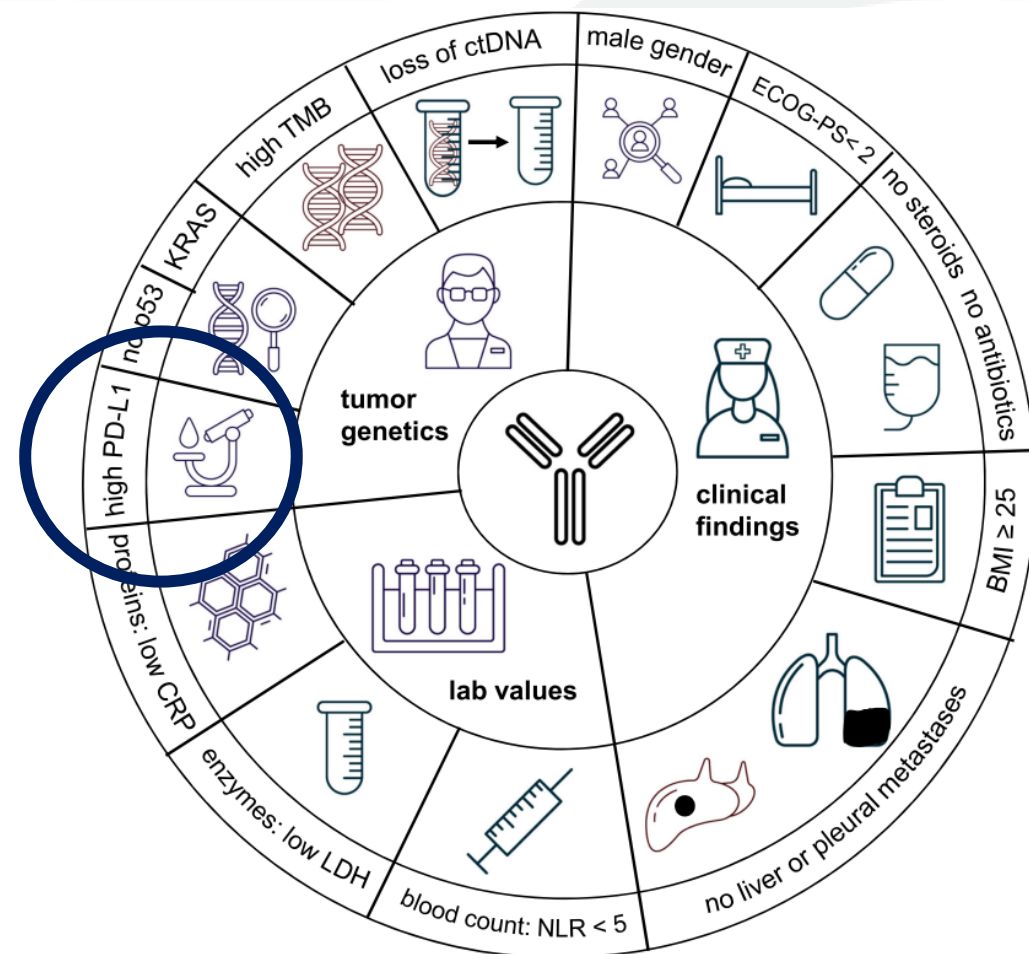
Mechanisms of resistance



PD-L1 testing

- Established predictive biomarker for immunotherapy (CPI targeting PD-1 & PD-L1)
- Higher the PD-L1 expression, higher the clinical ORR
15-25% response, ↑PFS and OS
- Testing technology is IHC
- Positivity defined with >1% PD-L1 expression on tumor cells

Positive predictive and prognostic factors for ICI therapy



Biomarker of Interest	Assay Details	Outcomes/Literature Support
Tissue-based		
PD-L1 Expression	Immunohistochemistry (IHC) to determine proportion of PD-L1 positivity/expression.	Greater PD-L1 positivity/expression associated with improved outcomes in first-line and second-line advanced NSCLC trials using IO. ¹³⁻¹⁵
Tumor Mutational Burden (TMB)	Whole exome sequencing or FoundationOne CDx assay to quantify the number of somatic mutations per coding area of a tumor genome.	Higher TMB associated with improved PFS, though not OS, with first-line ipilimumab/nivolumab in advanced NSCLC, irrespective of PD-L1 expression. ¹⁷
Tumor Infiltrating Lymphocytes (TILs)	Assessment of lymphocyte infiltration seen within tumor tissue.	Higher TIL density associated with improved survival in NSCLC. ^{47,48} Extent of PD-L1 expression on TILs associated with response to atezolizumab. ⁵⁰
Tumor Specific Genotypes	Fluorescence in situ hybridization (FISH) or next generation sequencing to identify genomic alterations in EGFR, ALK, KRAS etc.	EGFR and ALK mutated tumors associated with poorer outcomes in second-line IO trials. ⁵⁴ STK11/LKB1 co-mutation associated with IO resistance. ⁶¹
Gene Expression Signatures	Multi-gene profiling to identify immunogenic gene signatures, e.g. activated T-cell, IFN-γ	High expression of T-effector and INF-γ related gene signature associated with improved OS with second-line atezolizumab in advanced NSCLC. ²⁷
Serum-based		
Complete Blood Count (CBC) Markers (NLR, PLR, etc.)	Neutrophil to lymphocyte ratio (NLR), platelet to lymphocyte ratio (PLR), absolute eosinophil count, and others as calculated from CBC differential.	Higher NLR associated with poorer prognosis in advanced NSCLC. ^{71,72} NLR correlated to treatment response in second-line nivolumab studies. ^{74,75}
Blood Tumor Mutational Burden (bTMB)	FoundationOne CDx with quantification of single nucleotide variants, GuardantOMNI CDx assay.	Higher bTMB associated with longer PFS with second-line atezolizumab in advanced NSCLC. ⁷⁸ High bTMB subgroup with improved OS with first-line tremelimumab/durvalumab. ¹⁹

Neoadjuvant / perioperative treatment

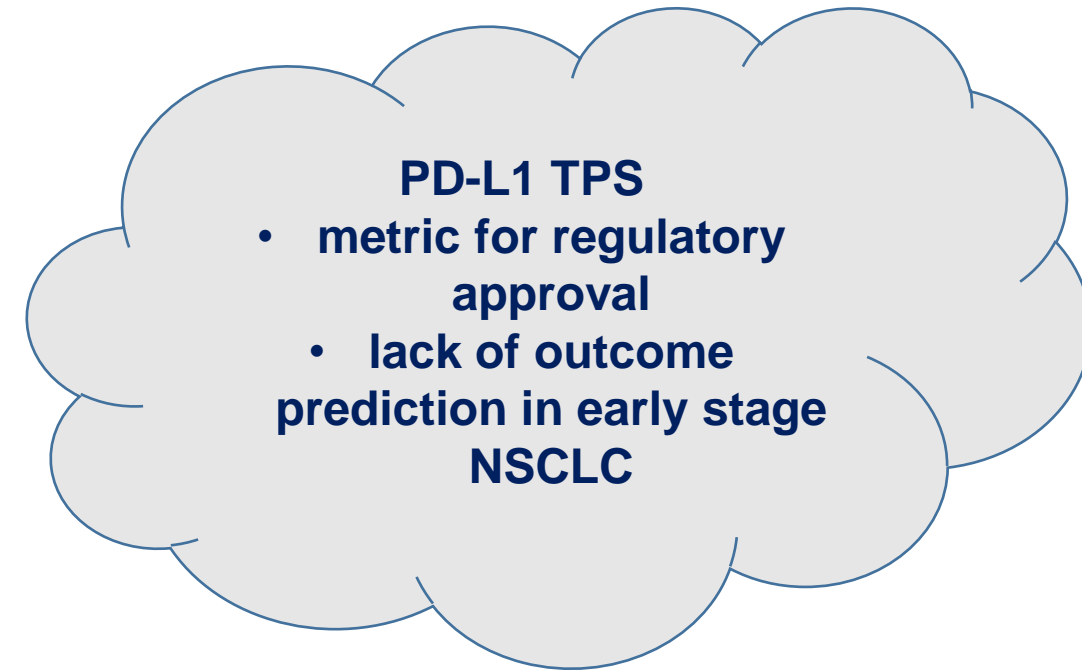
- NSCLC resectable
- Early stages (IIa – IIIb)
 - Tumor size > 4 cm
 - Tumor size < 4 cm + lymph node metastasis
- Chemotherapy + immunotherapy
- **TPS PD-L1 >1%*** → IHC
 - * EMA approval (nivolumab)
- **EGFR & ALK neg** → RT-PCR + IHC/FISH or RT-PCR or NGS



For patients being considered for neoadjuvant or adjuvant systemic therapy, at a minimum, determination of EGFR and ALK alteration status is required. Tumor proportion score measurement for determination of PD-L1 status should also be considered.

- Other biomarkers testing is encouraged → NGS

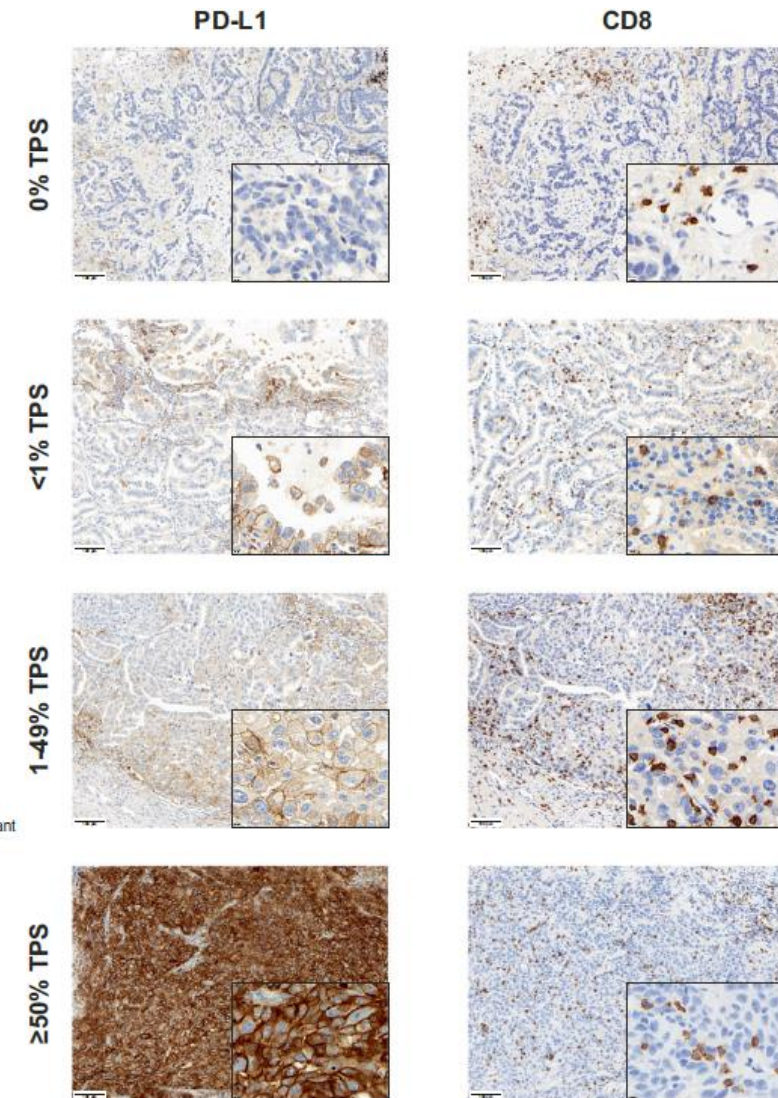
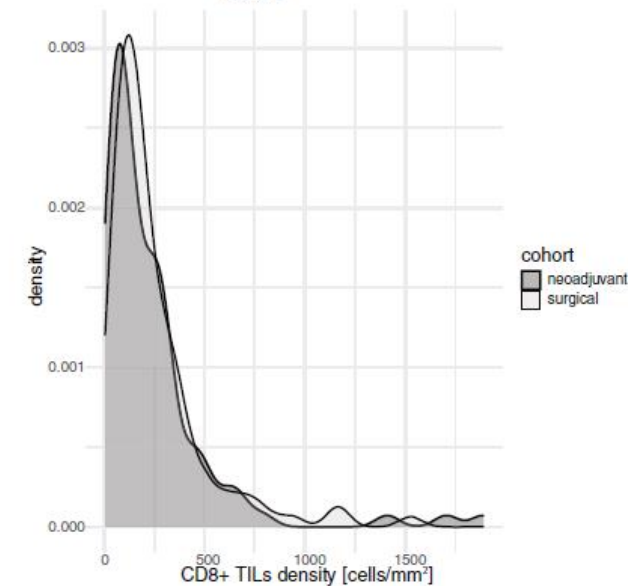
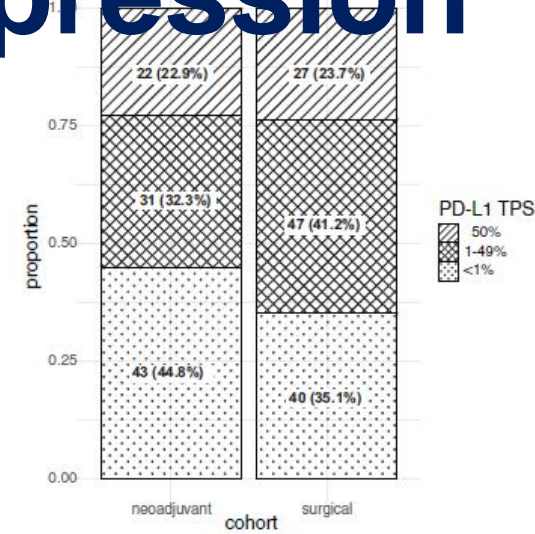
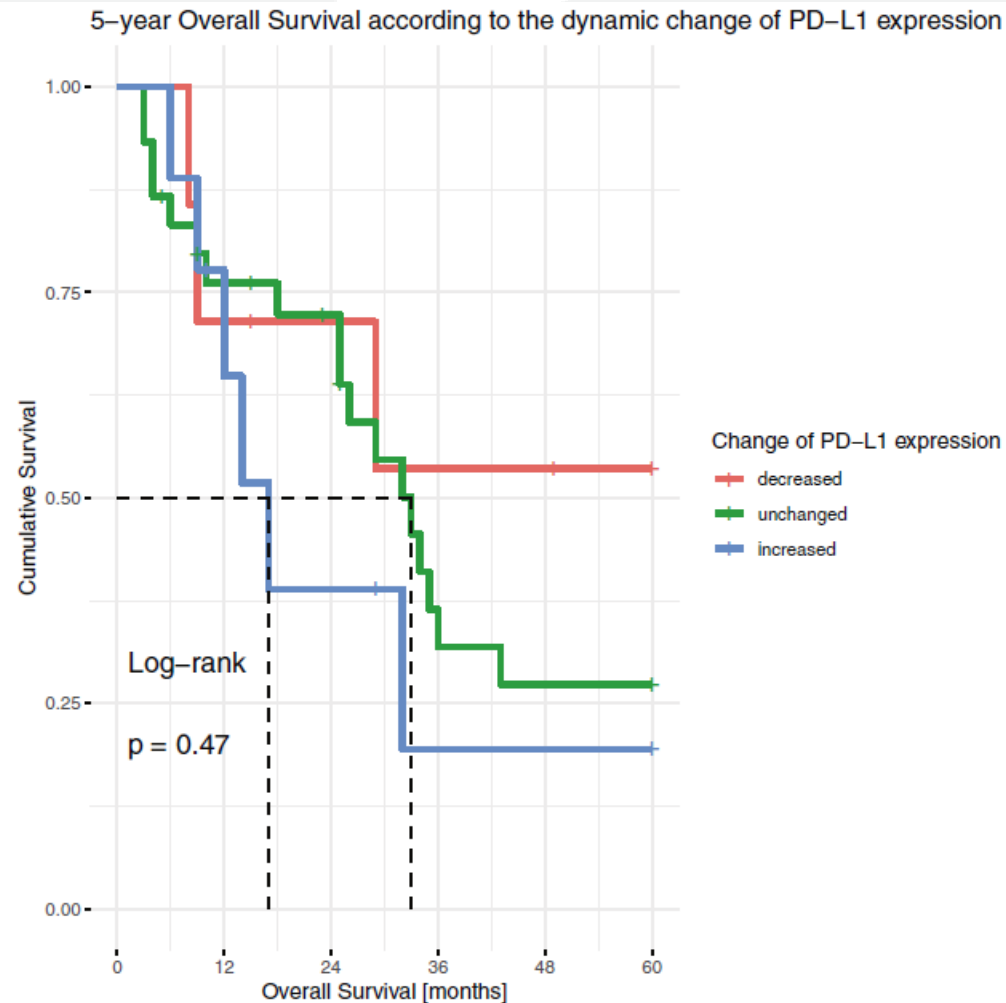
- Upfront /reflex in primary specimen
 - Should be considered
 - Accelerate time to treatment
 - Optimal lab processing logistics
- or/and
- Retesting in surgical specimen
 - For adjuvant treatment decision



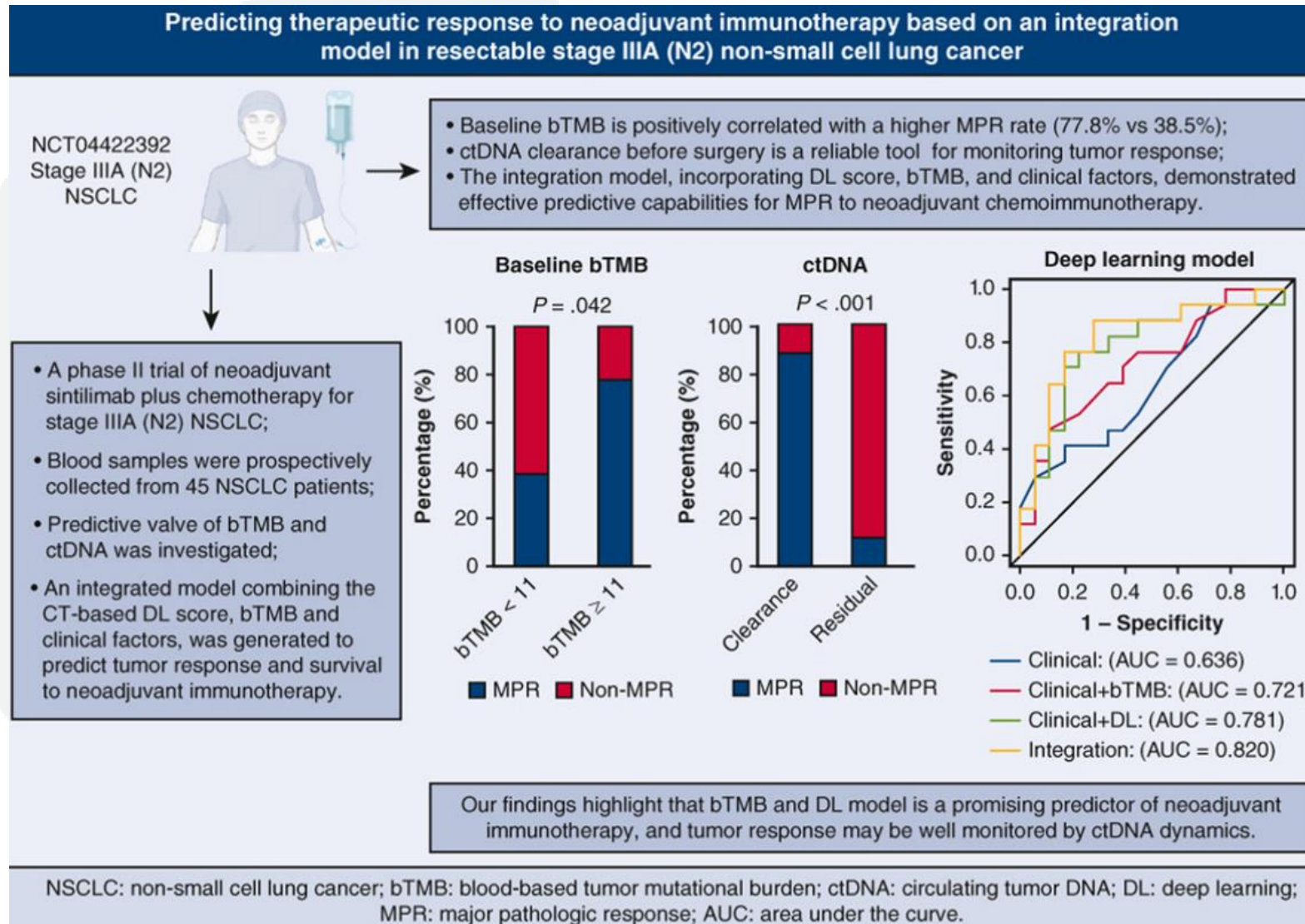
Patients with stage II or IIIA EGFR- and ALK-wild-type disease who have undergone complete resection followed by chemotherapy should be considered for adjuvant immunotherapy based on PD-L1 results as follows:

- PD-L1 < 1%: Discourage
- PD-L1 1%–49%: Consider
- PD-L1 > 50%: Recommend

Effect of neoadjuvant therapy on PD-L1 expression



TMB instead of PD-L1

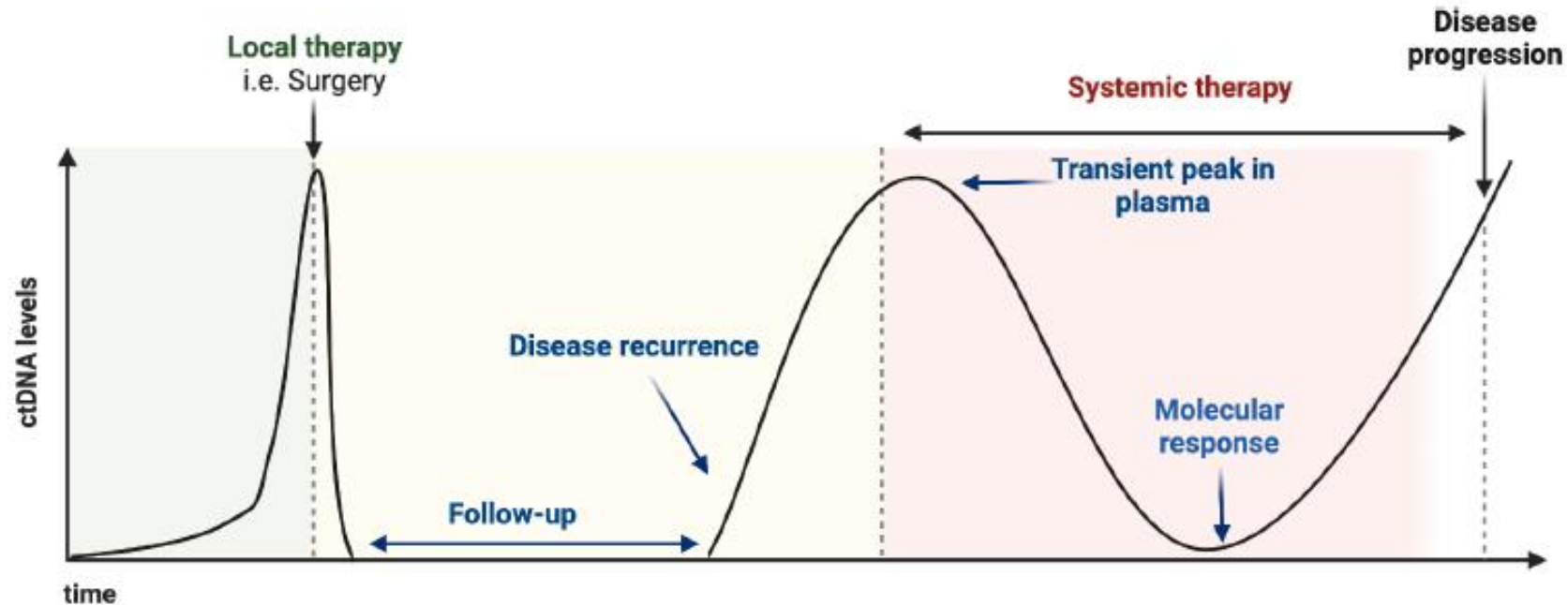


Programmed Death Ligand-1 and Tumor Mutation Burden Testing of Patients With Lung Cancer for Selection of Immune Checkpoint Inhibitor Therapies

Guideline From the College of American Pathologists, Association for Molecular Pathology, International Association for the Study of Lung Cancer, Pulmonary Pathology Society, and LUNGeity Foundation

Guideline Statement	Strength of Recommendation
1. In patients with advanced NSCLC, pathologists should use a validated PD-L1 IHC expression assay, in conjunction with other targetable genomic biomarker assays where appropriate, to optimize selection for treatment with ICIs.	Strong recommendation
2. Pathologists should ensure appropriate validation has been performed on all specimen types and fixatives. <i>Note:</i> Specific validation requirements are out of the scope of this guideline, and laboratories should refer to the Principles of Analytic Validation of Immunohistochemical Assays Guideline ⁵⁷ for details on how to validate IHC specimens.	Conditional recommendation
3. When feasible, pathologists should use clinically validated PD-L1 IHC assays as intended.	Conditional recommendation
4. Pathologists who choose to use LDTs for PD-L1 expression should validate according to the requirements of their accrediting body.	Strong recommendation
5. Pathologists should report PD-L1 IHC results using a percentage expression score.	Conditional recommendation
6. Clinicians should not use tumor mutation burden alone to select patients with advanced NSCLC for ICIs, based on insufficient evidence in this population.	Conditional recommendation

Liquid biopsy



Applications of
ctDNA analyses

Screening and
Early detection

MRD monitoring

Tumor
genotyping
for therapy
selection

Treatment monitoring

Tumor
genotyping
for the
identification
of resistance
mechanisms

Liquid biopsy in NSCLC advanced stage

Sequential steps

A *



1. Tissue biopsy first



2. Liquid biopsy second

B



1. Liquid biopsy first



2. Tissue biopsy second

Simultaneous steps

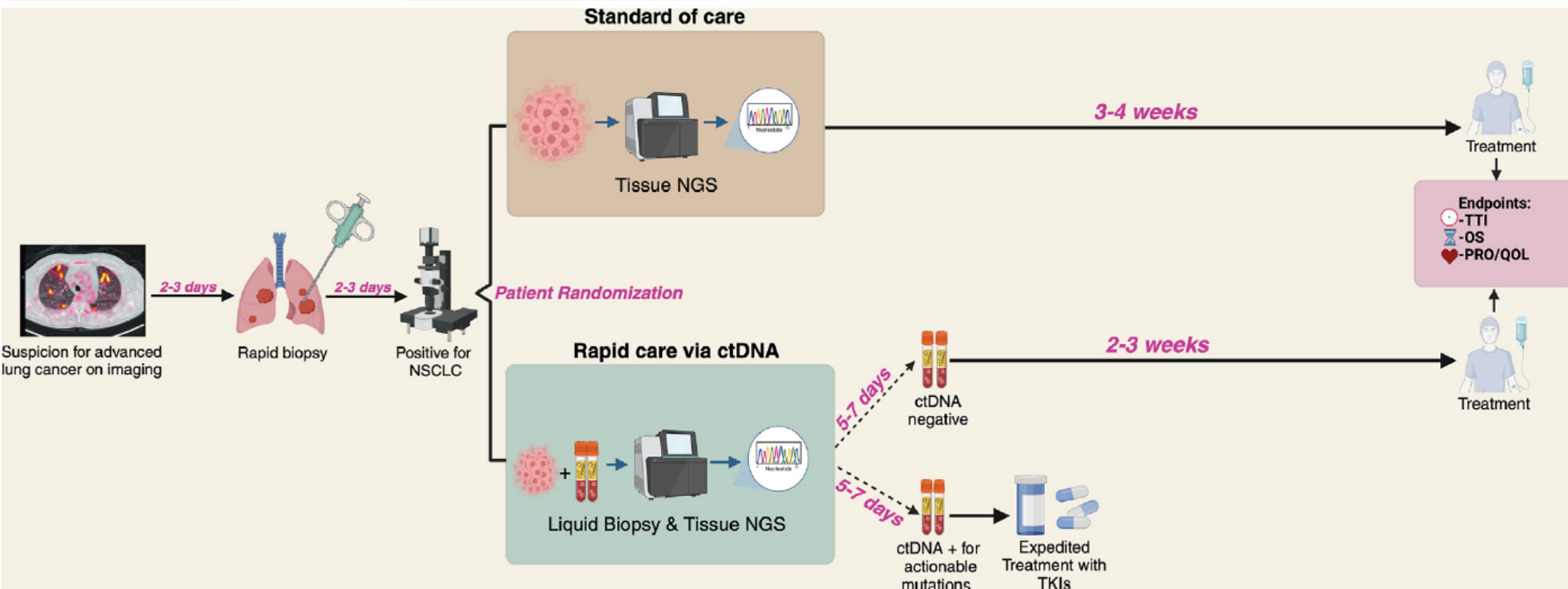
C



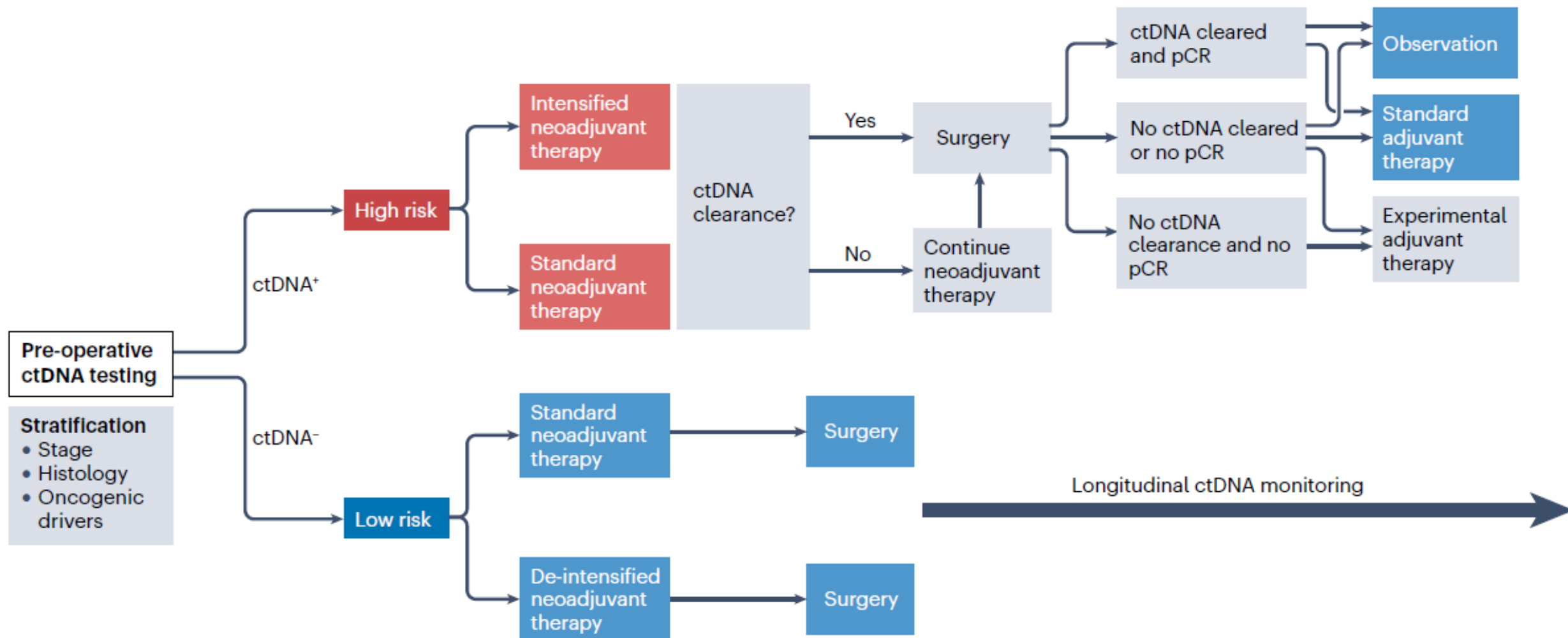
Tissue and liquid biopsies

* According to the International guidelines (from the IASLC and ESMO)

Shorter time to treatment with liquid biopsy



ctDNA in neoadjuvant setting



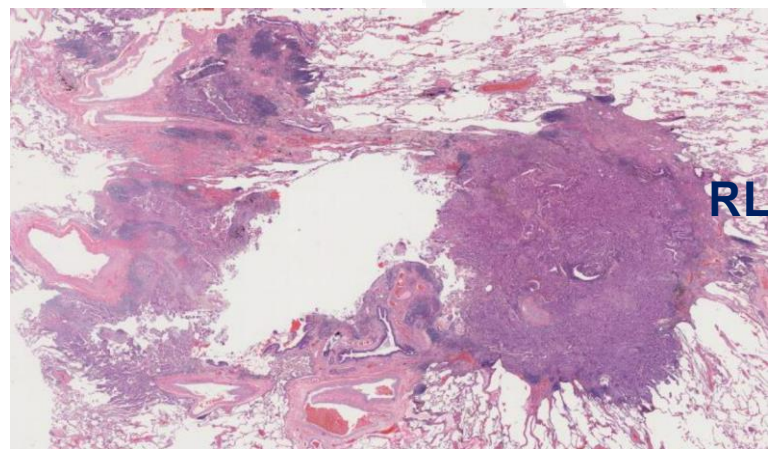
Multiple lung tumors



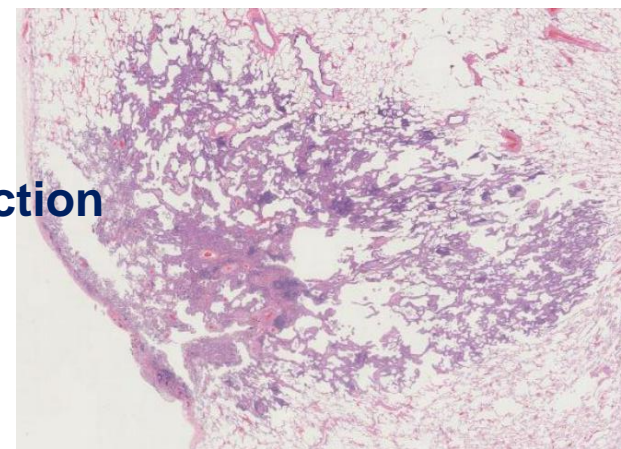
- D = 15 mm
- ADC G2
- Point mutation RAF1, p.S259T, VAF 5,1 %



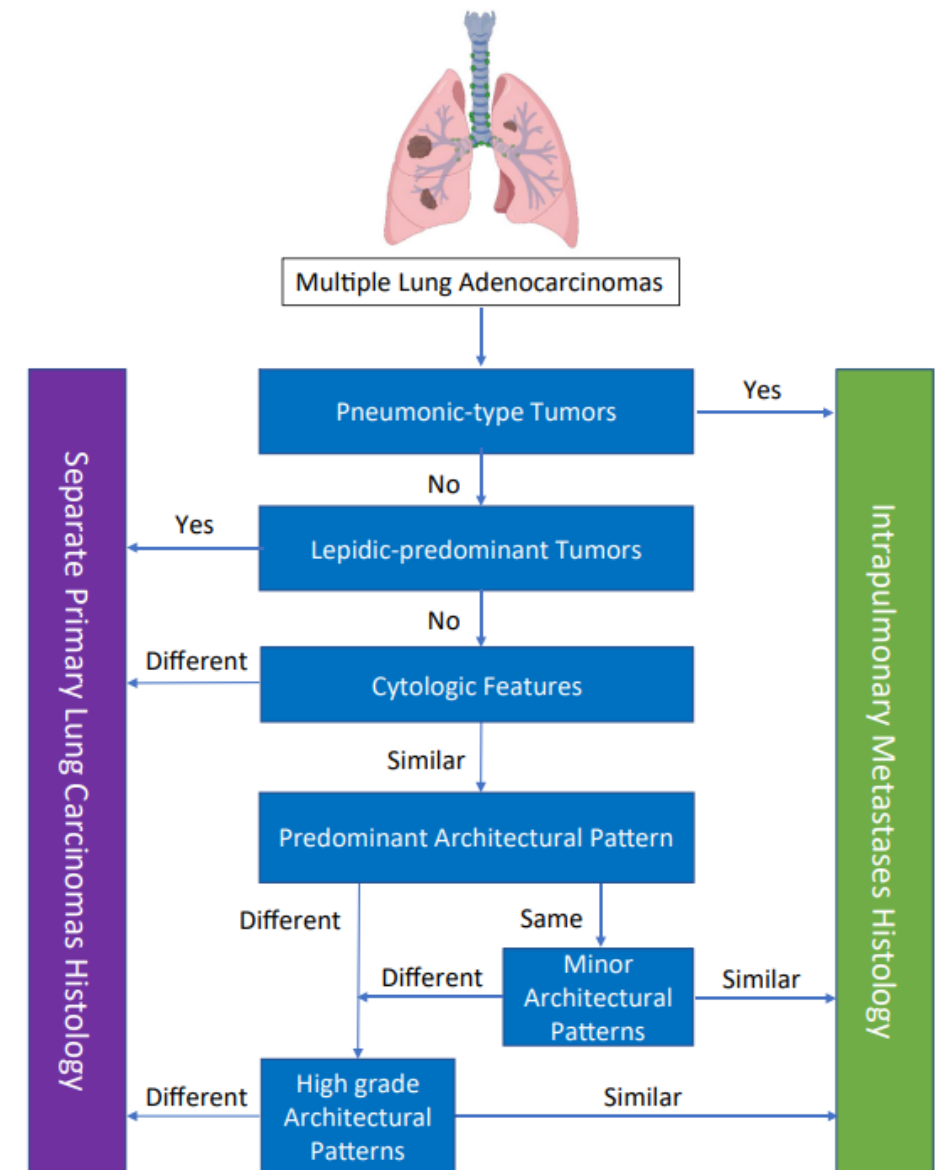
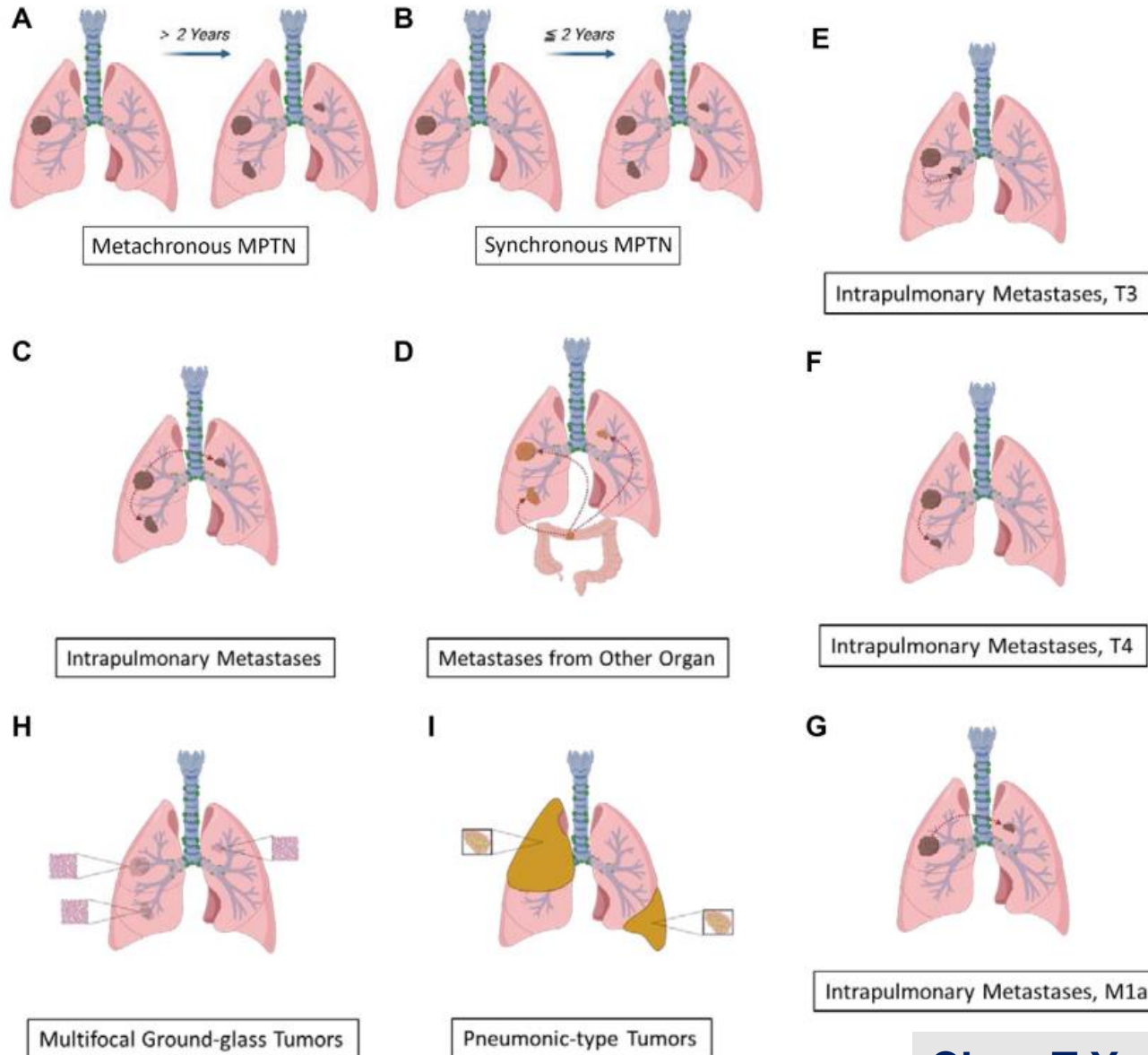
- D = 14 mm
- MIA
- Point mutation KRAS, p.G12V, VAF 15,5 %



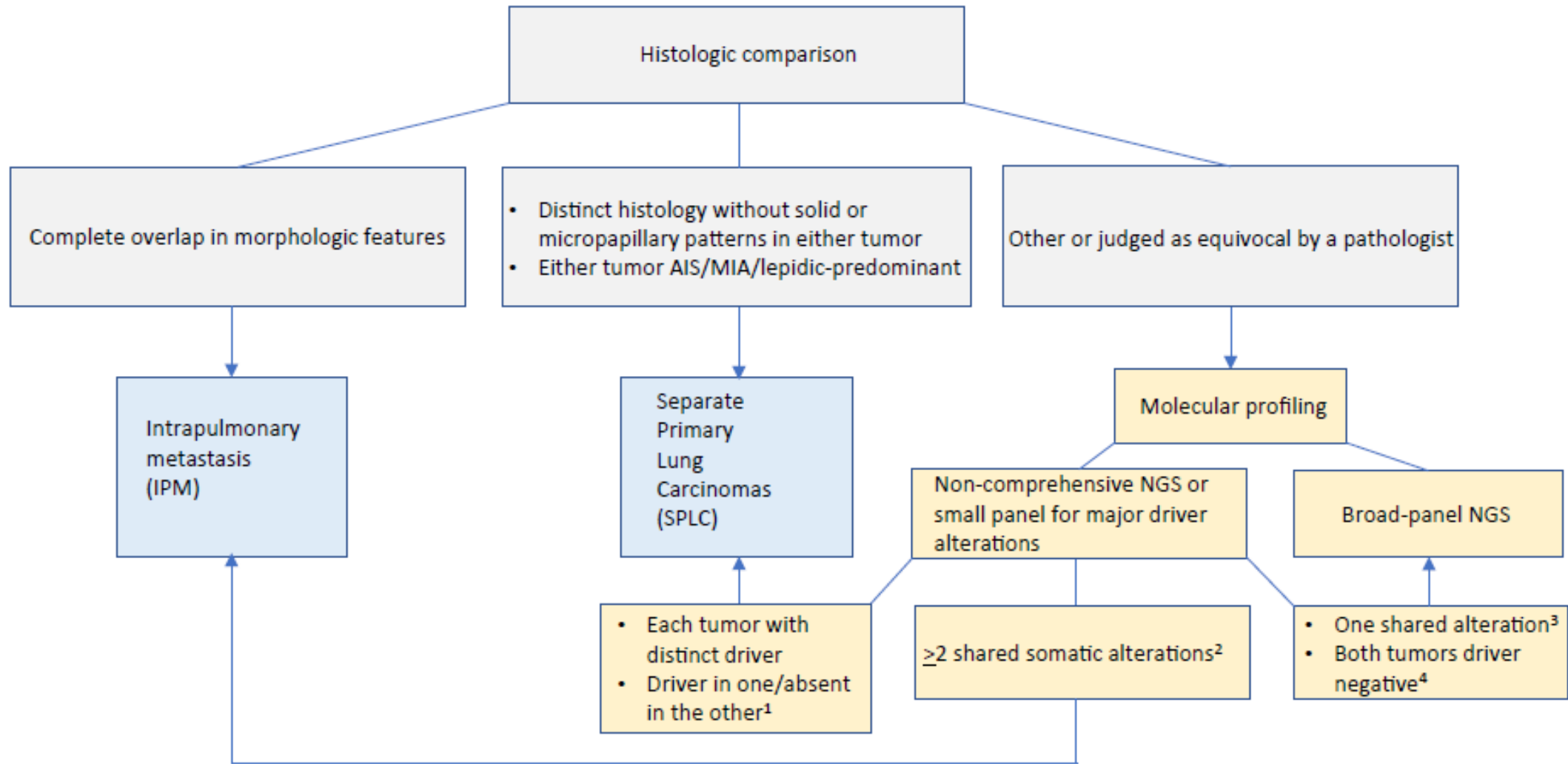
RLL lobectomy + RUL wedge resection



Primary LADC vs IPM



Multiple NSCLC

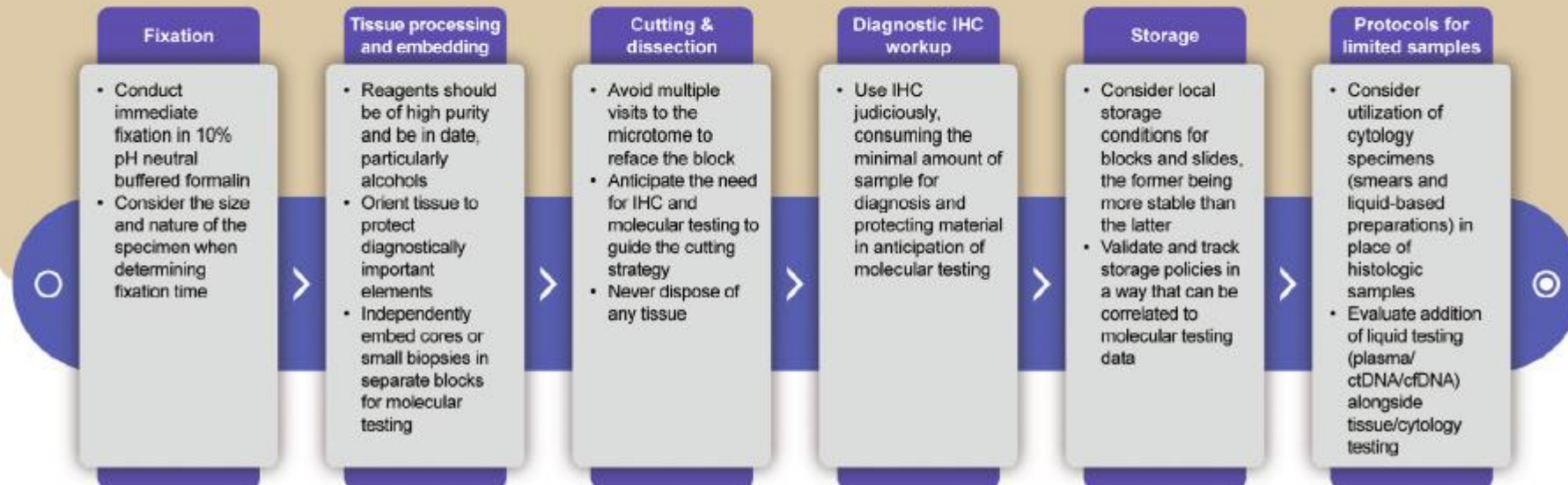


Optimizing tissue path

Overarching considerations

- From when lung cancer is first suspected, all stakeholders must consider that predictive biomarker testing will likely be required, which, in turn, should drive practices and procedures to conserve tissue appropriately.
- The availability of, and adherence to, clear, robustly validated laboratory procedures must be ensured. This extends to appropriate staff training, tracking appropriate metrics, and taking corrective or preventive actions as required.
- Laboratory staff must be well versed in the optimal handling of different sample types.
- Communication channels between pathologists, laboratory colleagues, external laboratories, and other members of the MDT must be open, bidirectional, and routine.

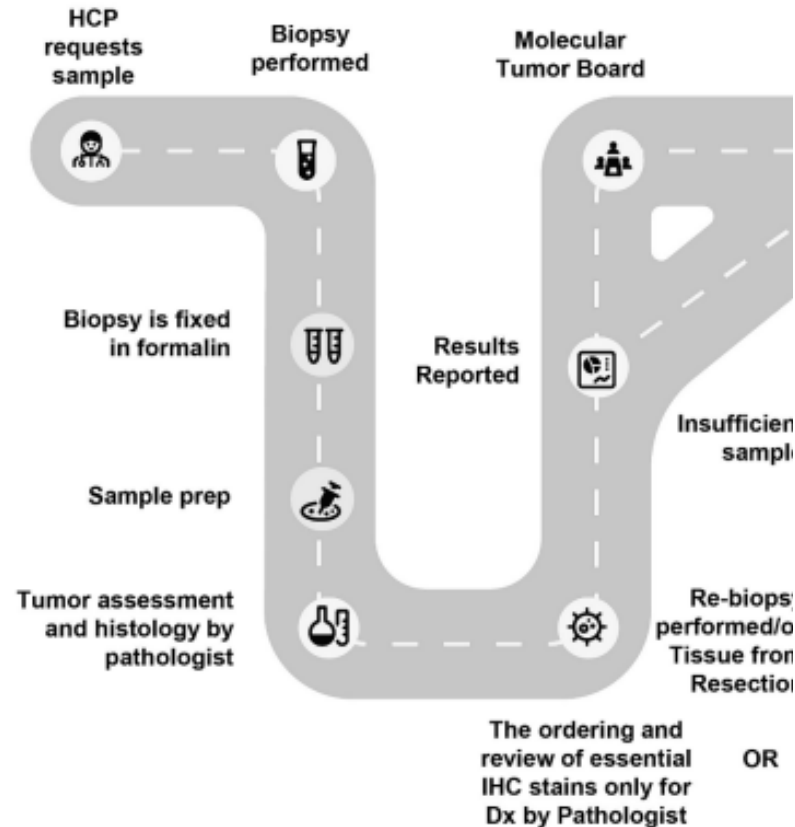
Process-specific guidance



Predictive biomarker testing implementation



Continued education of staff on the advancement



Challenge	Recommendations
Biomarker testing in operable NSCLC is not routinely requested	<ul style="list-style-type: none"> - More frequent revision and harmonization of guidelines across various markets and regions. - Education of pathologists and physicians and laboratories to ensure they are aware of newly available therapies and associated testing. - Regulations to ensure that reimbursement for pertinent biomarker testing becomes available simultaneously with the approval of targeted drugs.
Insufficient tissue for biomarker testing from the initial diagnostic biopsy	<ul style="list-style-type: none"> - Ensuring that tissue conserving practices are followed at diagnosis to allow sufficient material for downstream biomarker testing. - Utilisation of alternative sample types such as cytology or liquid biopsies when tissue is not available.
Biomarkers are tested for individually instead of in within a multigene NGS panel	<ul style="list-style-type: none"> - Implementation of frameworks (upscaling of capacity, quality assurance, availability of reimbursement) to allow access to NGS testing. - Formation of multi-disciplinary virtual molecular tumour boards (MTBs) for interpreting complex data to guide treatment decisions.

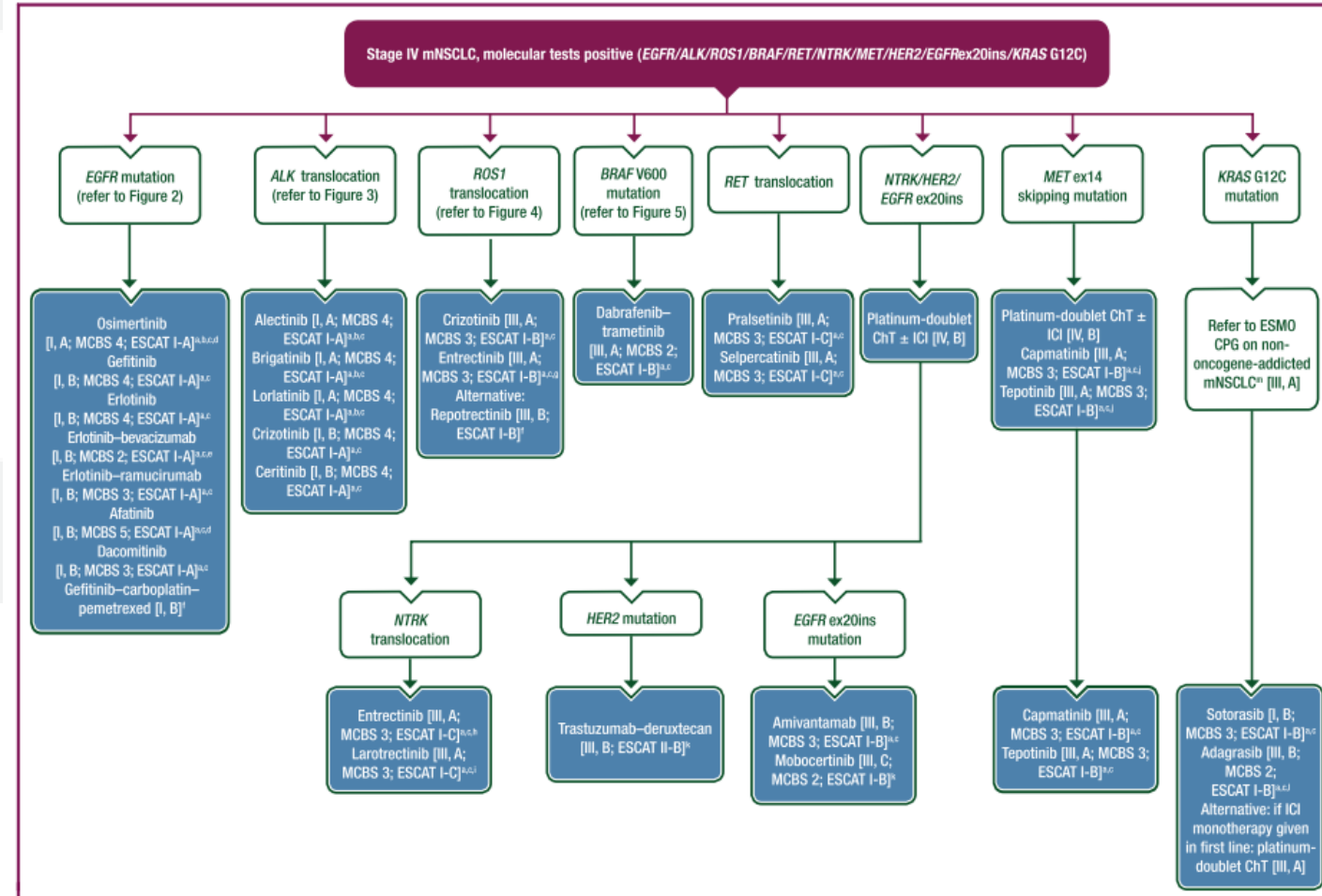
Guidelines & recommendations - molecular



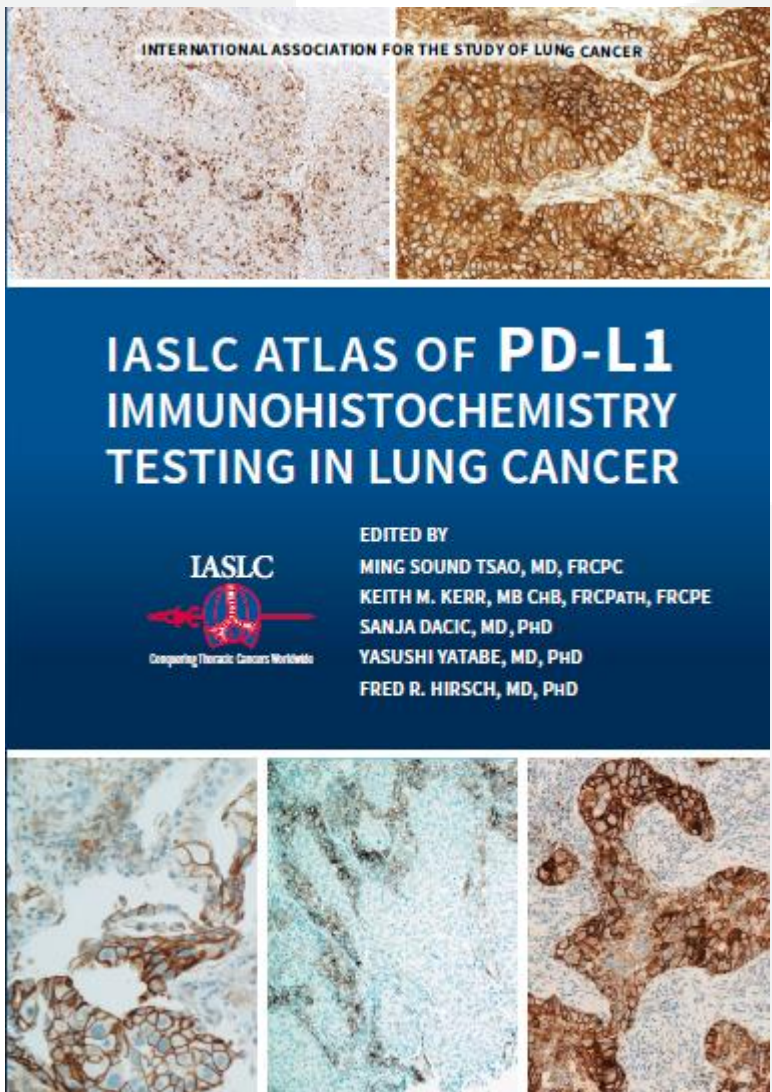
IASLC ATLAS OF MOLECULAR TESTING FOR TARGETED THERAPY IN LUNG CANCER

ESMO guidelines 2023

Newly diagnosed patients	EGFR	Any validated method to cover mutations in exon 18-21 (DNA NGS preferred)
	ALK	RNA NGS; IHC ± molecular confirmation (NGS, FISH)
	ROS1	RNA NGS; IHC screening, molecular confirmation essential (NGS, FISH)
	RET, MET, NTRK, ERBB2 (HER2), KRAS, BRAF	DNA/RNA NGS panel testing
	PD-L1	IHC
	EGFR T790M, MET (as appropriate) (cfDNA/tissue DNA)	PCR/NGS/ISH
Relapsed patients on targeted therapy	EGFR (category 1)	Broad molecular profiling (NGS) ^c



Guidelines & recommendations – PD-L1



- PD-L1 IHC should be systematically determined in advanced NSCLC [I, A]

PD-L1 expression	IHC to identify PD-L1 expression at the appropriate level and on the appropriate cell population(s) as determined by the intended drug and line of therapy. Only specific trial assays are validated. Internal and external quality assurance are essential	To enrich for those patients more likely to benefit from anti-PD-1 or anti-PD-L1 therapy. For pembrolizumab, testing is a companion diagnostic for nivolumab and atezolizumab, testing is complementary	I, A
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ESMO guidelines 2019

REVIEW ARTICLE

PD-L1 Testing for Lung Cancer in 2019: Perspective
From the IASLC Pathology Committee

JTO 2019

testing for PDL1 is now recommended
for advanced-stage non-neuroendocrine carcinomas



Never ending story

Variable	ctDNA [91–93]	miRNA [94–96]	bTMB [54,81,97,98]	Immunological Markers [20,99,100]
Type of Biomarker	Genetic (circulating DNA)	Genetic (non-coding RNA)	Genetic (mutational burden)	Protein (immune proteins)
Detection Method	NGS, digital PCR	Real-time PCR, microarrays	NGS, digital PCR	IHC, flow cytometry
Clinical Utility	Diagnosis, prognosis, monitoring	Prognosis, monitoring	Prognostic, predictive	Diagnostic, predictive
Prognostic and Predictive Aspects	High sensitivity for early detection	Correlates with immunotherapy response	Predicts response to specific immunotherapies	Expression correlated with survival and response
Variability Factors	Influenced by tumor burden, detection techniques	Influenced by sample conditions	Requires standardization in measurement	Sensitive to detection methods and immune status
Advantages	Non-invasive, high sensitivity	Non-invasive, easily quantifiable	Information on tumor heterogeneity	Directly related to mechanisms of action of therapies
Limitations	Cost, need for sequencing	Inter- and intra-individual variability	Influenced by technical and biological factors	Requires validation for specific interpretation
Cost-Effectiveness	Moderate–high	Low–moderate	High due to sequencing technologies	Moderate, depends on the marker and method
Usage Recommendations	Widely recommended in clinical guidelines	In research, some clinical applications	Recommended in specific contexts	Emerging use, supported by recent studies
Recent Innovations	Advances in digital PCR technology	New miRNAs associated with NSCLC	Improvements in accuracy and cost of NGS	New predictive markers of response to PD-1/PD-L1